



# Continuum Diffusion Rate of Enzymes by Solving the Smoluchowski Equation

CHEM 276 & BENG 275

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[http://mccammon.ucsd.edu/smol/doc/tutorials/chem276\\_smol.pdf](http://mccammon.ucsd.edu/smol/doc/tutorials/chem276_smol.pdf)

# Objectives

- Basic theories of Poisson-Boltzmann and Smoluchowski Equations.
- Experiment 1: the born ion
- Experiment 2: mouse AChE enzyme
- Assignments

# Lecture Review

- Before you start this tutorial, please try to answer the below questions:
  1. What's the Poisson-Boltzmann equation (PBE)? What does it describe? Why do we need to solve it and what's the output?
  2. What's the Smoluchowski diffusion equation? What's the relationship with the PBE? What's the problem domain? Is it the same with the PBE?

# Poisson-Boltzmann equation

$$-\nabla \cdot \varepsilon(x) \nabla U(x) + \bar{\kappa}^2(x) \sinh U(x) = \frac{4\pi e_c^2}{kT} \sum_i z_i \delta(x - x_i)$$

Linearized Poisson-Boltzmann equation also useful:

$$-\nabla \cdot \varepsilon(x) \nabla U(x) + \bar{\kappa}^2(x) U(x) = \frac{4\pi e_c^2}{kT} \sum_i z_i \delta(x - x_i)$$

$$U(x)|_{x \in \partial\Omega} = g(x)|_{x \in \partial\Omega}$$

Additional notation for charge distribution term:

$$f(x) = \frac{4\pi e_c^2}{kT} \sum_i z_i \delta(x - x_i)$$

# Smoluchowski Equation

Describes the over-damped diffusion dynamics of non-interacting particles in a potential field.

$$\frac{\partial p(\vec{r}, t | \vec{r}_0, t_0)}{\partial t} = -\nabla \cdot D \left[ \nabla - \beta \vec{F}(\vec{r}) \right] p(\vec{r}, t | \vec{r}_0, t_0)$$

Or for  $\vec{F}(\vec{r}) = -\nabla U(\vec{r})$

$$\begin{aligned} \frac{\partial p(\vec{r}, t | \vec{r}_0, t_0)}{\partial t} &= -\nabla \cdot J p(\vec{r}, t | \vec{r}_0, t_0) \\ &= -\nabla \cdot D e^{-\beta U(\vec{r})} \nabla e^{\beta U(\vec{r})} p(\vec{r}, t | \vec{r}_0, t_0) \end{aligned}$$

# Steady-state Formation

$$\frac{\partial p(\vec{r}, t)}{\partial t} = 0$$

$$\Rightarrow \nabla \cdot D e^{-\beta U(\vec{r})} \nabla e^{\beta U(\vec{r})} p(\vec{r}, t | \vec{r}_0, t_0) = 0$$

Suppose

$$D'(\vec{r}) = D e^{-\beta U(\vec{r})} \quad \text{and} \quad p'(\vec{r}, t) = e^{\beta U(\vec{r})} p(\vec{r}, t | \vec{r}_0, t_0)$$

Finally, we have

$$-\nabla \cdot D'(\vec{r}) \nabla p'(\vec{r}, t) = 0$$

$$\longrightarrow p(\vec{r}, t | \vec{r}_0, t_0) = e^{-\beta U(\vec{r})} p'(\vec{r}, t)$$

# Modeling procedure

1. According the above couple of slides, we have to solve PBE using APBS first and then read the output potential into the Smoluchowski equation to solve the diffusion equation.
2. Today we try to finish Exp. 1, if you have interest, you can continue Exp. 2.
3. Ask your tutor for help to accomplish this tutorial. For further questions about the diffusion solver, please go to SMOL homepage:

<http://mccammon.ucsd.edu/smol>

4. All the tutorial materiers can be downloaded from here:

# Tutorial directory guide

- **NOTE:**

```
tar vczf smol.tar.gz
```

```
cd $HOME/smol
```

```
source ./tcshrc
```

- **Data structures**

./bin /\* all the executable binary files \*/

./tcshrc /\* set some necessary environments \*/

./mesh /\* all the mesh files we will use for this tutorial \*/

./pqr /\* all the PQR files for this tutorial \*/

./potential /\*all the potential scripts for APBS runs. \*/

./run /\* You can do your work under this directory. \*/

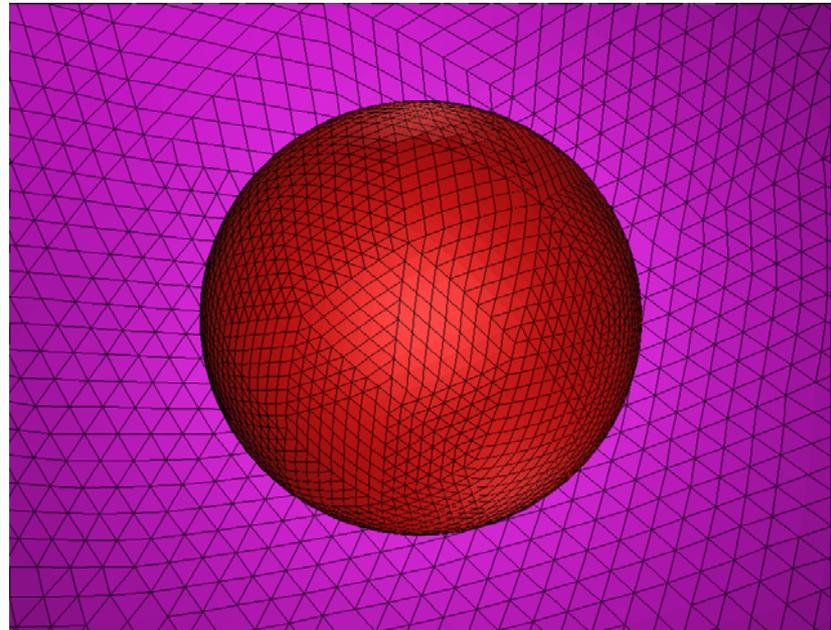
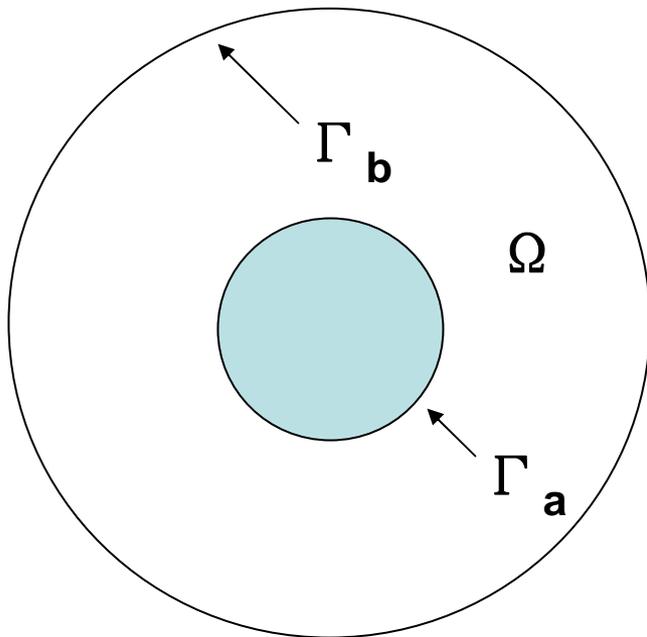
./tools /\* Here are some visualization scripts. \*/

# Exp. 1: Mesh preparation

Mesh preparation: Netgen 4.4 (<http://www.hpfem.jku.at/netgen/>)

Netgen is an excellent mesh generator, especially for the spherical shaped objects.

The finite problem domain is the spherical test case.



# Exp. 1: Simple mesh generation

Our first task is to generate the analytical test for the SMOL diffusion.

software: Netgen (<http://www.hpfem.jku.at/netgen/>)

software tutorial: (<http://www.hpfem.jku.at/netgen/ng4.pdf>)

```
source $HOME/smol/tcshrc
```

```
cd $HOME/smol/mesh/born
```

```
ng
```

Start from “file”, then “Load Geometry”, then “Generate Mesh”.

Note: The node and element numbers are shown below the software screen

Then you can refine the mesh by choosing “Refinement”. (For example, I have stored a case with 409,886 vertices. )

Finally, from “file”->“Export Mesh”, save the mesh as “born.mesh”

# Exp. 1: Simple mesh generation

```
cp born.mesh mesh.neu
```

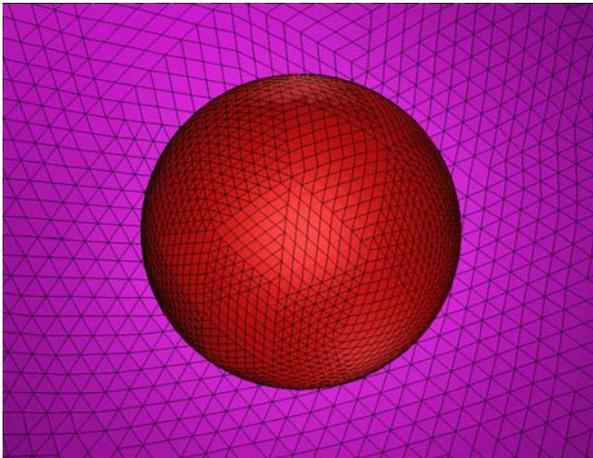
```
neu2m >& born.m
```

born.m is exactly the input file we will use for this tutoring.

To visualize your mesh, you can type

```
mcsf2off --boundary born.m
```

```
geomview born.off (OR mcsf born.off)
```



# Exp. 1: analytical solution

For a spherically symmetric system with a Coulombic form of the PMF,  $W(r) = q/(4\pi\epsilon r)$ , the SSSE can be written as

$$\frac{1}{r^2} \frac{\partial}{\partial r} (r^2 Jp) = \frac{1}{r^2} \frac{\partial}{\partial r} \left( r^2 D \left( \frac{\partial p}{\partial r} - \beta p \frac{qq_l}{4\pi\epsilon r^2} \right) \right) = 0$$

Suppose  $Q = \frac{\beta qq_l}{4\pi\epsilon}$ ,  $p(r_1) = 0$ ;  $p(r_2) = p_{bulk}$

Then,  $k_{on} = \frac{4\pi Q D r_1^2}{\frac{-Q}{e^{r_2}} - \frac{-Q}{e^{r_1}}}$       If  $Q = 0$ ,  $k_{on} = \frac{4\pi D r_1^2}{\frac{1}{r_1} - \frac{1}{r_2}}$

# SMOL sample input files

- **NOTE:**

- # model parameters
- charge 0.0 /\* ligand charge \*/
- conc 1.0 /\* initial ligand concentration at the outer boundary \*/
- diff 78000.0 /\* diffusion coefficient \*/
- temp 300.0 /\* temperature, unit: Kelvin \*/
- # potential gradient methods
- METHtype FEM /\* you can choose BEM or FEM \*/
- # mapping method
- map DIRECT /\* you can choose NONE/DIRECT/FEM \*/
- # steady-state or time-dependent
- tmkey SSSE /\* you can choose SSSE or TDSE \*/
- # input paths
- mol ../../pqr/ion\_yuhui.pqr
- mesh ../../mesh/sphere\_4.m
- mgrid ../../potential/pot-0.dx /\* for APBS input \*/
- dPMF ../../force/evosphere\_4.dat /\* for BEM input \*/
- end 0

# Manage your input parameters

- **NOTE:**

`#{solver}`

- the default input file: `smol.in`

`#{solver} -ifnam filename`

- the default iteration method: `CG(lkey=2)`.

`BCG (lkey=4 or 5), BCGSTAB(lkey=6)`

`#{solver} -lkey 2`

- default maximal number of iterative steps: 5000

`#{solver} -lmax 8000`

# Manage your input parameters (cont.)

- **NOTE:**

- the default timestep:  $5.0 \cdot 10^{-6} \mu\text{s}$

`{solver} -dt 5.0*10-5`

- the default number of time steps: 500

`{solver} -nstep 1000`

- the default concentration output frequency: 50

`{solver} -cfreq 100`

- the default reactive integral output frequency: 1

`{solver} -efreq 5`

- the default restart file writing frequency: 1000

`{solver} -pfreq 5000`

# Exp. 1: Steady-state Diffusion calculation

```
cd $HOME/smol/run/born
```

```
vi solve-all.csh
```

- Please use any text editor to edit “solve-all.csh” to control your calculations.
- AND check your “smol-template.in”, you can use the potential files you calculated. Make sure that the potential path is correct.
- `./solve-all.csh >& ./solve-all.log &`

# Exp. 1: Steady-state Diffusion Output

```
cd $HOME/smol/run/born
```

- In “rate.\*.dat” file there are the  $k_{on}$  simulation and analytical values.

```
vi rate.*.dat
```

# Exp. 1: Visualization of your calculation

OpenDX is applied to show concentration distribution at steady state.

Please select some tutorials from the below list if you want to know more about OpenDX:

<http://ivc.tamu.edu/docs/opendx.pdf>

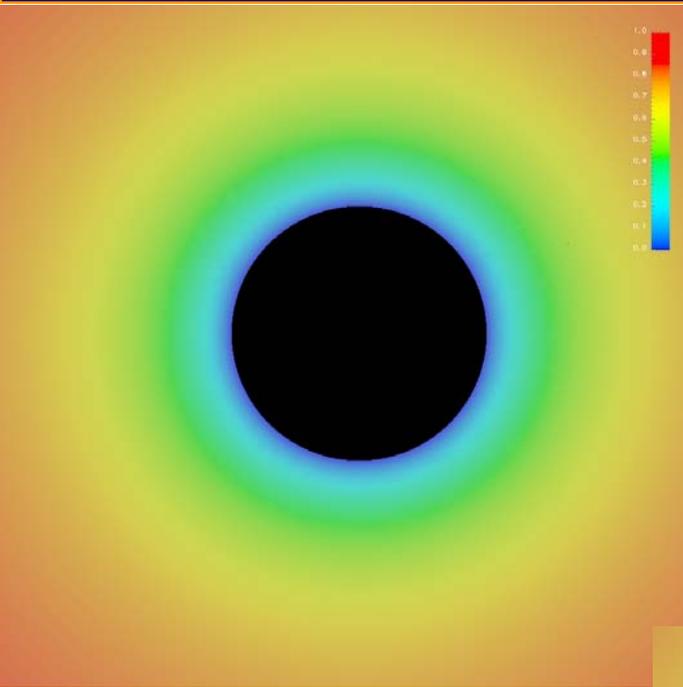
```
source $HOME/smol/tcshrc
```

```
cd $HOME/smol/run/born/anal.*.*
```

```
dx -edit ../../../../tools/visualization/conc.net
```

Please let your tutor know if you don't know how to use OpenDX and really want to learn.

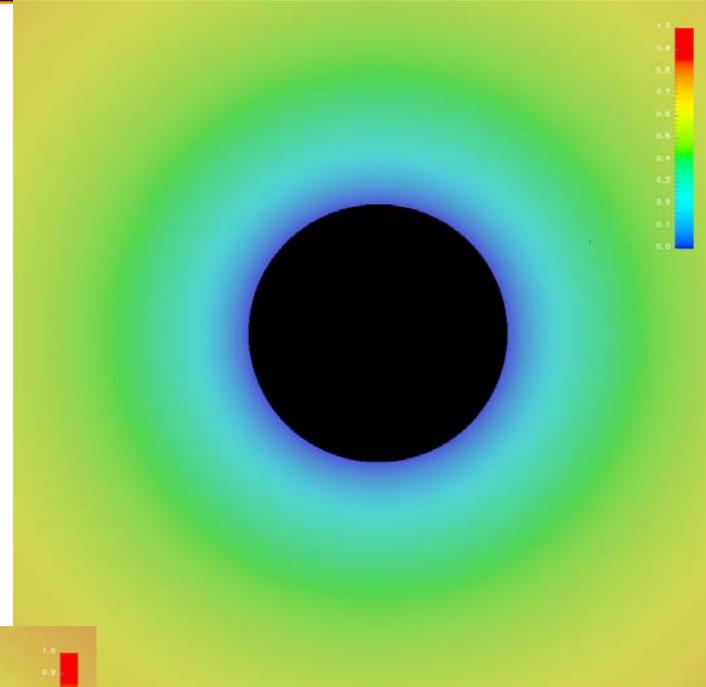
# Exp. 1: Sample output figures



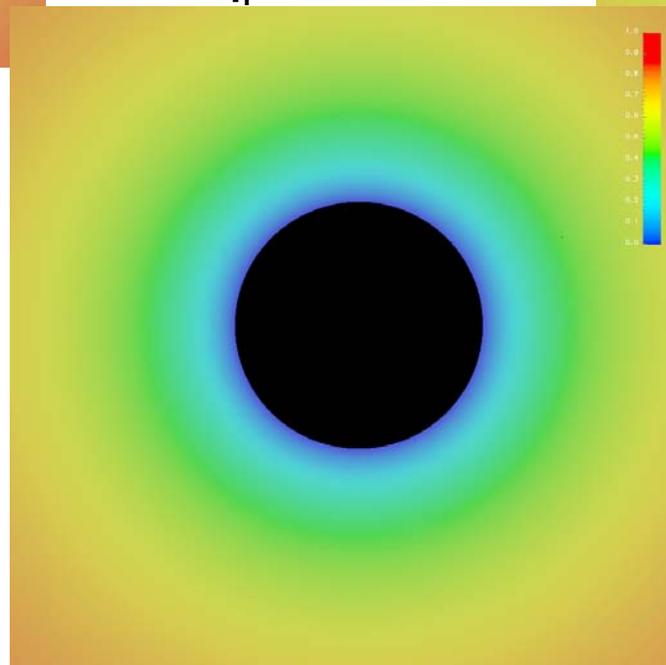
$$q_l = -1.0e$$

$q_l$  is the  
charge of the  
ligand.

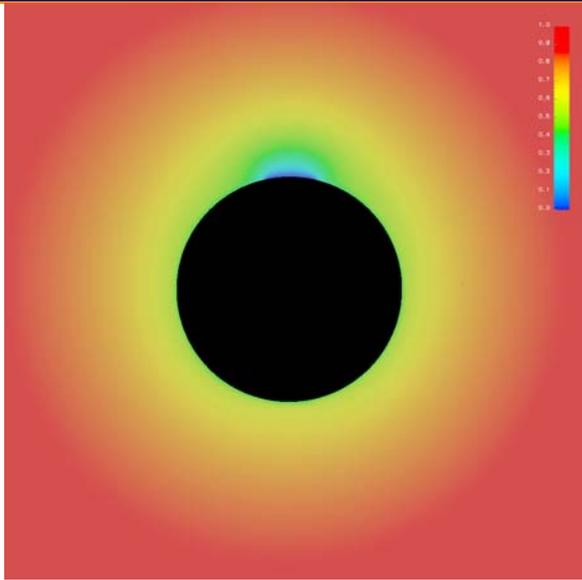
$$q_l = 0.0e$$



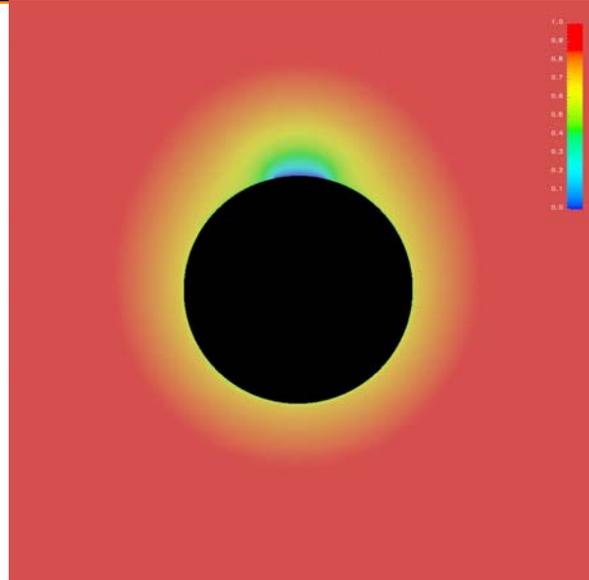
$$q_l = 1.0e$$



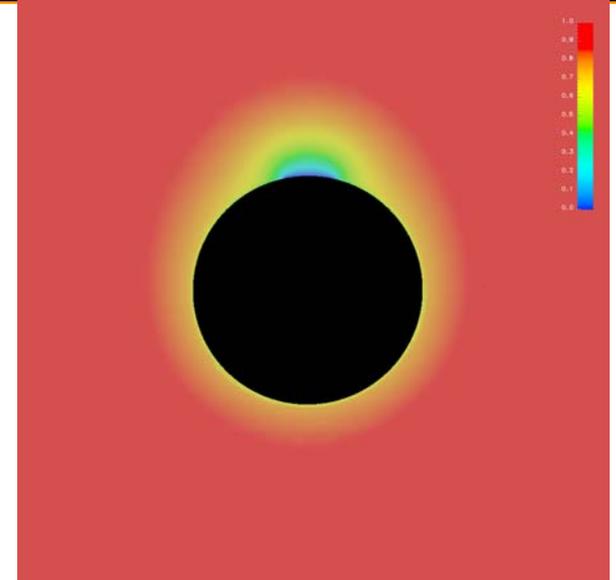
# Exp. 1: Sample output I ( $qq_1 = 1.0$ )



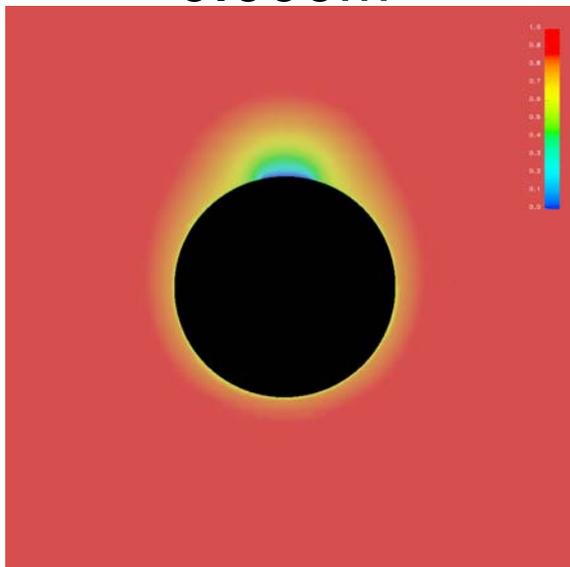
0.000M



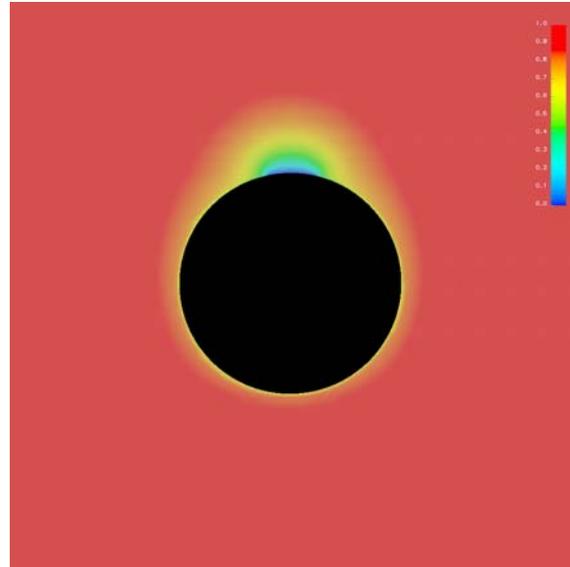
0.075M



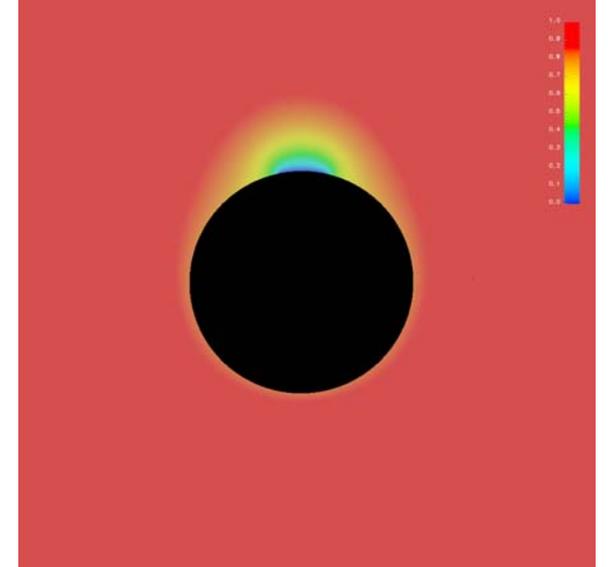
0.150M



0.300M

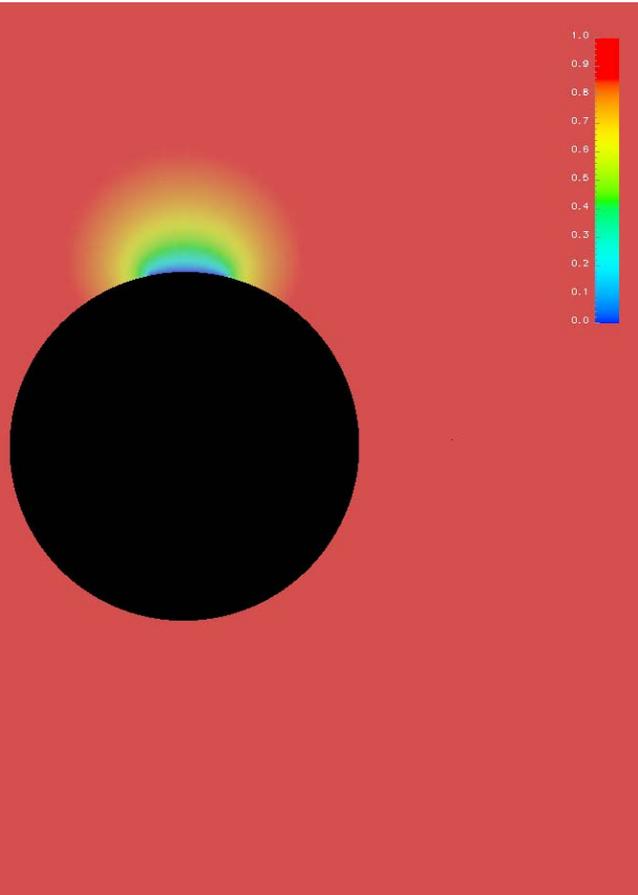


0.450M

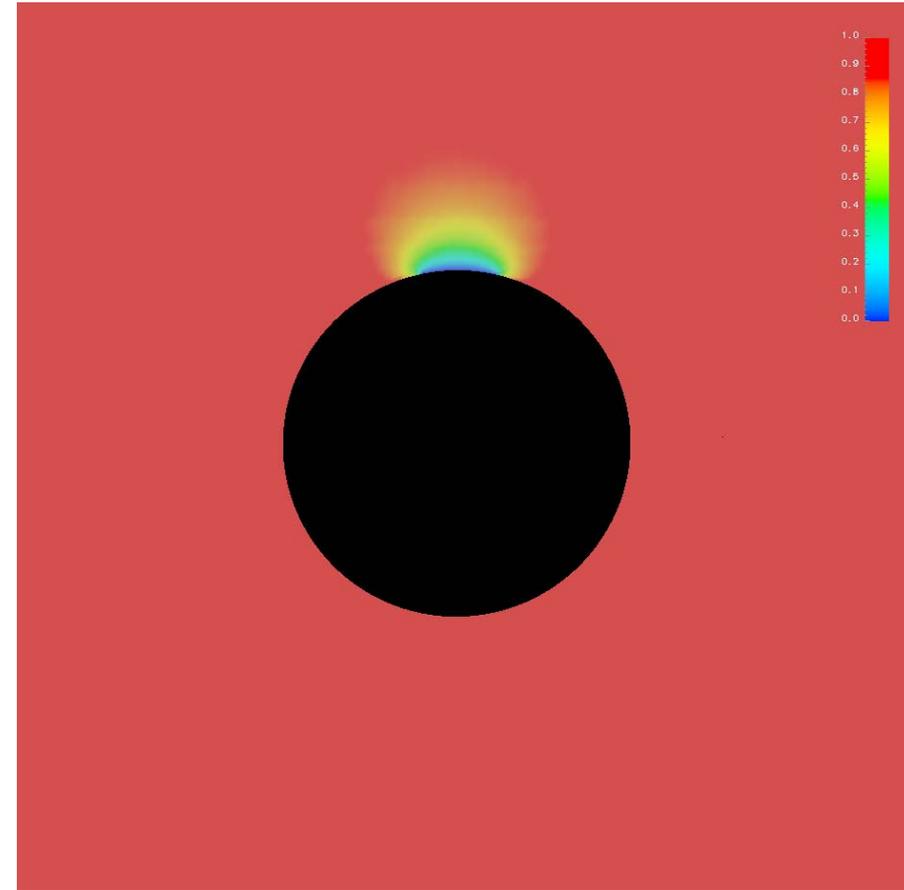


0.670M

# Exp. 1: Sample output II ( $qq_i = 0.0$ )



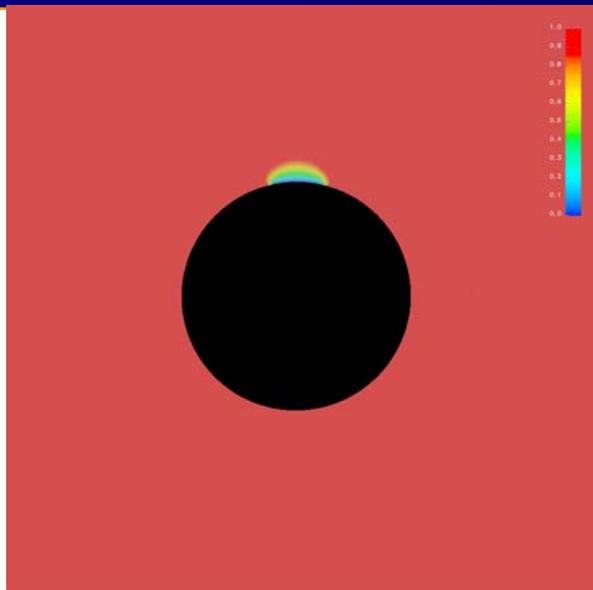
0.000M



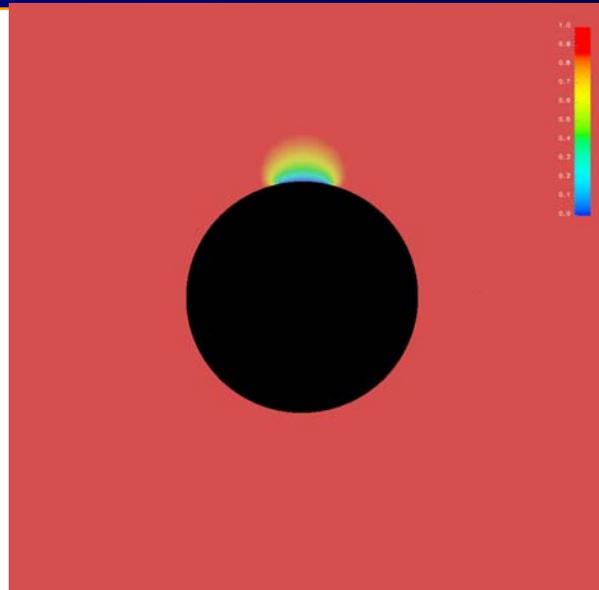
0.600M

Certainly, there is no difference at any ionic strength.

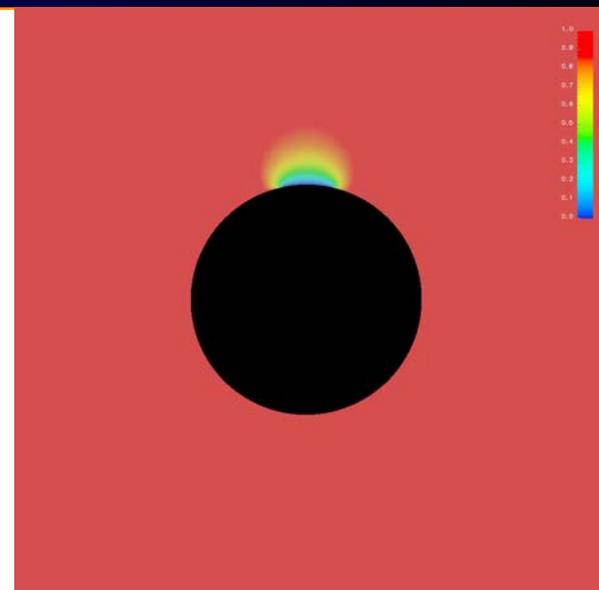
# Exp. 1: Sample output III ( $qq_1 = -1.0$ )



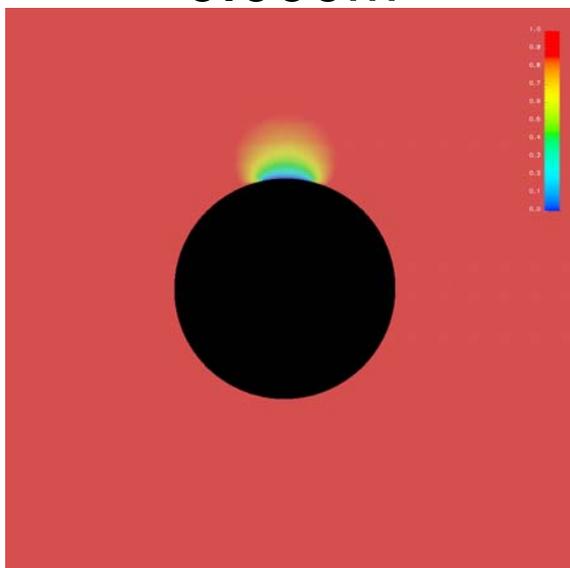
0.000M



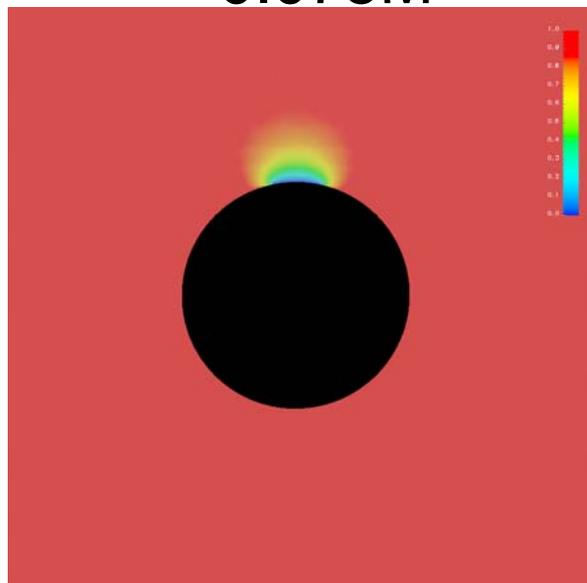
0.075M



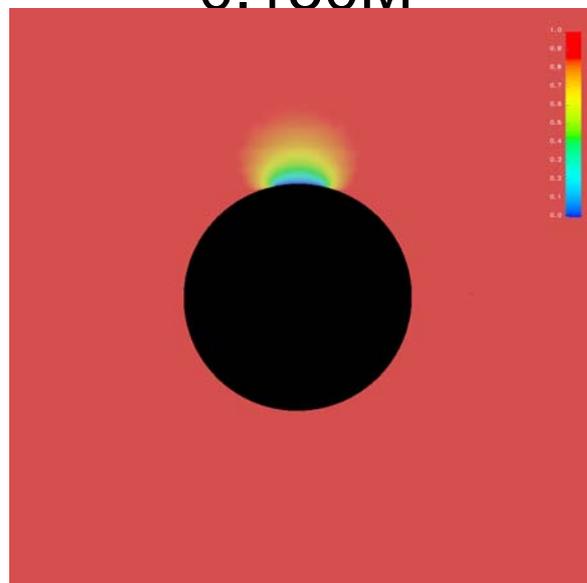
0.150M



0.300M



0.450M



0.600M

## Note

Your output should be different from the above three figures, for the whole molecular surface is active. However, part of the molecular surface has been assigned as the reactive boundary. Can you find which part of the molecule is reactive?

To learn how to assign the reactive boundary, please go through the next example: mouse acetylcholinesterase.

## Exp. 2: mesh and pqr file

➤ The mACHe mesh file was generated by Mol-LIBIE invented by Chandrajit's group.

➤ PQR file can be generated from PDB by Nathan's PDB2PQR server:

<http://pdb2pqr.sourceforge.net/>

➤ Assign the reactive boundary

Make sure to set the coordinate of carbonyl carbon of S203 at (0, 0, 0), and align the active site gorge with the y axis.

```
source $HOME/smol/tcshrc
```

```
cd $HOME/smol/mesh/mache
```

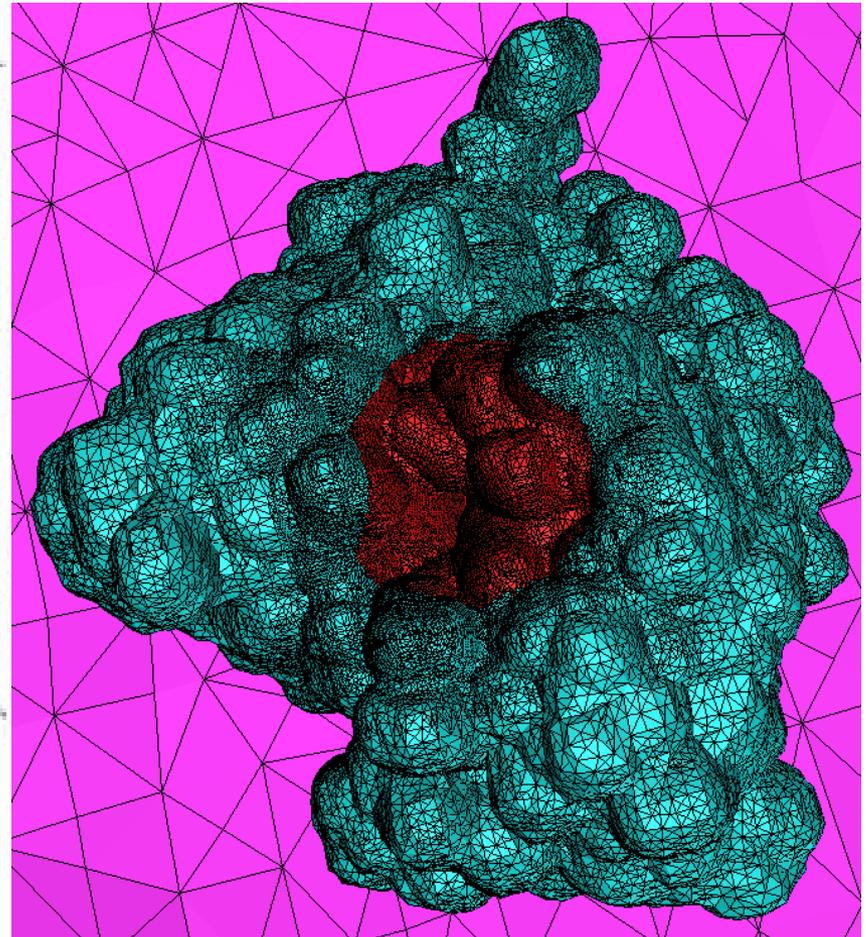
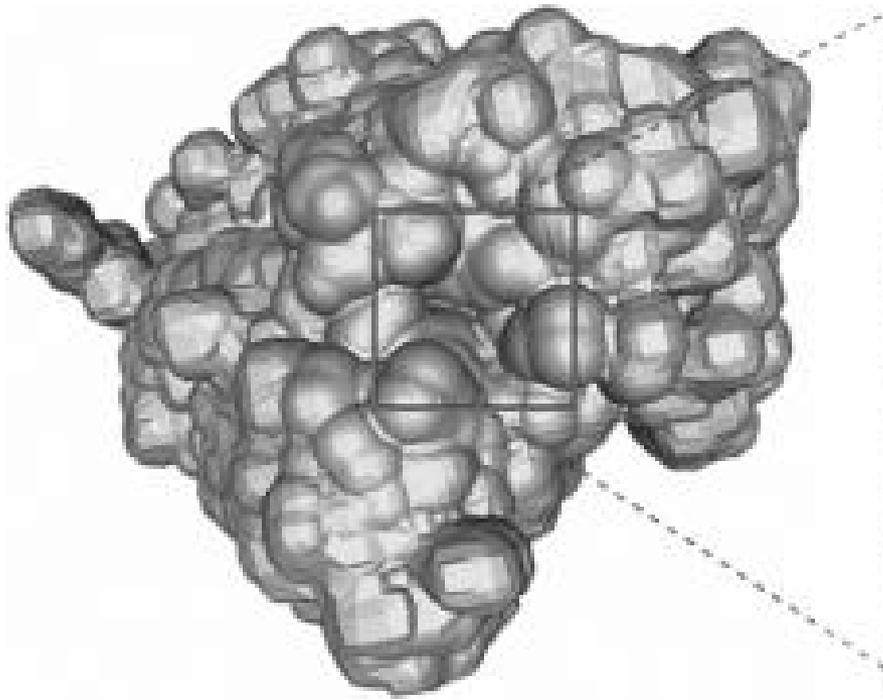
```
./assignBoundary.csh >& ./assignBoundary.log &
```

## Exp. 2: Visualize your new mesh

To visualize your mesh, you can type

```
mcsf2off --boundary mol-bc1.m
```

```
geomview mol-bc1.off (OR mcsf mol-bc1.off)
```



# Exp. 2: potential calculation

## Note:

1. I cannot guarantee the below calculation can successfully be done using APBS, since it might need big memory and large quota of the hard drive. Please ask your tutor for help if you undergo any trouble.

2. You can edit the value of “i” in “calc-all-pot.csh” to execute different calculations.

```
source $HOME/smol/tcshrc
```

```
./ calc-all-pot.csh >& calc-all-pot.log &
```

# Exp. 2: steady-state diffusion calculation

```
source $HOME/smol/tcshrc  
cd $HOME/smol/run/mache  
./solve-all.csh >& solve-all.log &
```

Please be patient to wait a couple of minutes to read output...

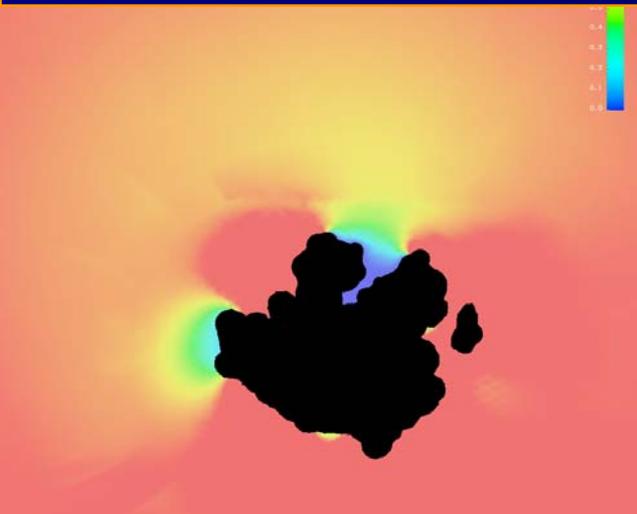
# Exp. 2: Visualization of your calculation

```
source $HOME/smol/tcshrc
```

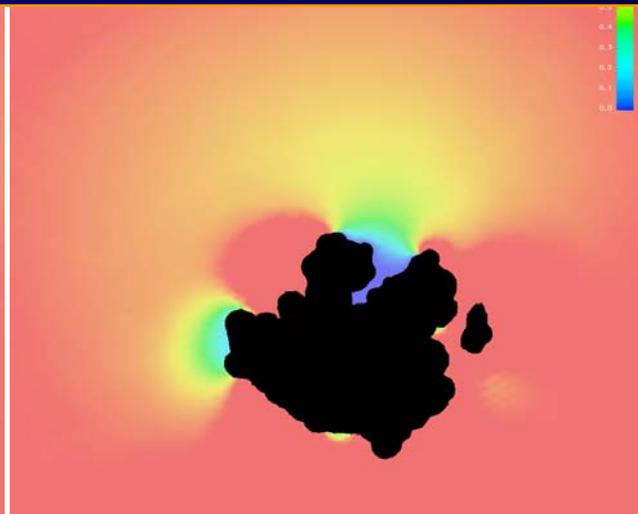
```
cd $HOME/smol/run/mache/mache.*.*
```

```
dx -edit ../../../../tools/visualization/conc.net
```

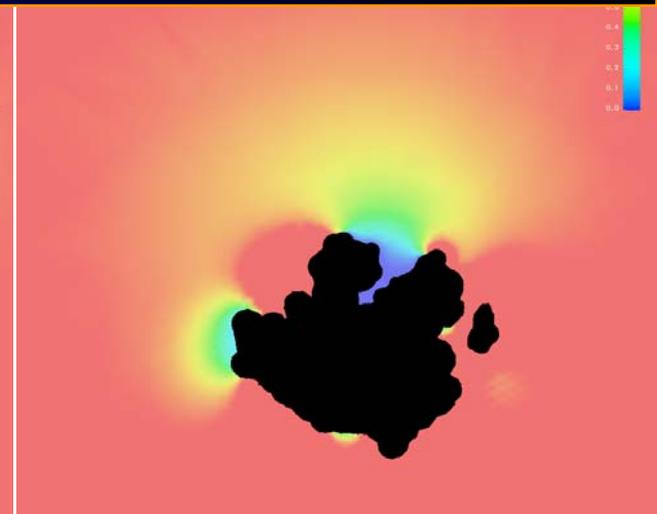
# Exp. 2: Sample outputs



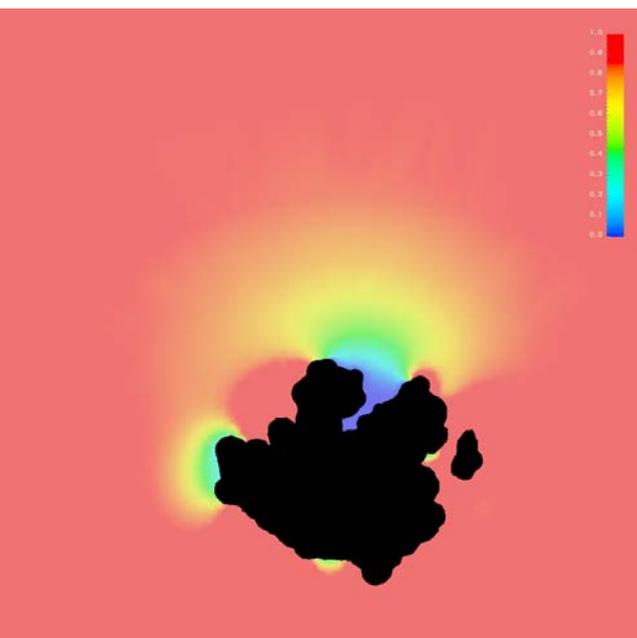
0.025 M



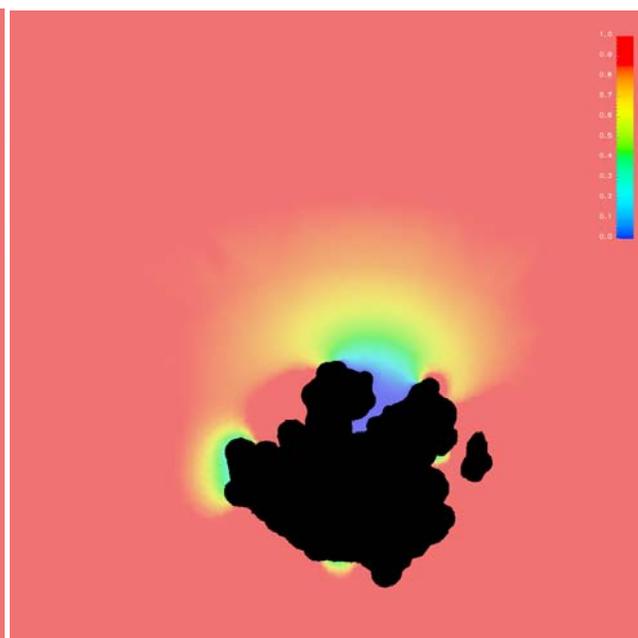
0.050 M



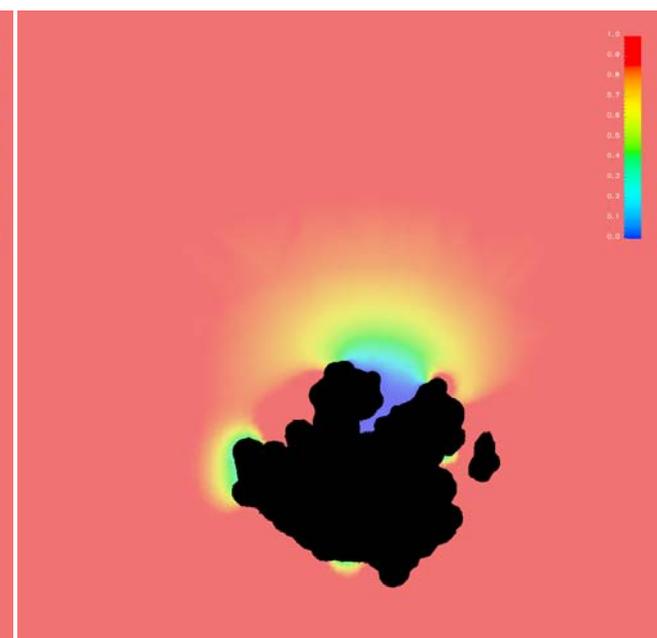
0.100 M



0.225 M



0.450 M



0.670 M

# Additional reading materials

1. <http://en.wikipedia.org/wiki/Diffusion>
2. Berg, H C. *Random Walks in Biology*. Princeton: Princeton Univ. Press, 1993
3. advanced diffusion materials:  
<http://www.ks.uiuc.edu/Services/Class/PHYS498NSM/>
4. Adaptive Multilevel Finite Element Solution of the Poisson-Boltzmann Equation I: Algorithms and Examples. *J. Comput. Chem.*, 21 (2000), pp. 1319-1342.
5. Finite Element Solution of the Steady-State Smoluchowski Equation for Rate Constant Calculations. *Biophysical Journal*, 86 (2004), pp. 2017-2029.
6. Continuum Diffusion Reaction Rate Calculations of Wild-Type and Mutant Mouse Acetylcholinesterase: Adaptive Finite Element Analysis. *Biophysical Journal*, 87 (2004), pp.1558-1566.
7. Tetrameric Mouse Acetylcholinesterase: Continuum Diffusion Rate Calculations by Solving the Steady-State Smoluchowski Equation Using Finite Element Methods. *Biophysical Journal*, 88 (2005), pp. 1659-1665.
8. Finite Element Analysis of the Time-Dependent Smoluchowski Equation for Acetylcholinesterase Reaction Rate Calculations. *Biophysical Journal*, 92(2007), 3397-3406

# Assignments

1. Please modify the keyword “SSSE” in “smol-template.in” to “TDSE”, i.e. to solve time-dependent SMOL equation instead of steady-state SMOL equation, rerun the whole scripts, what will happen?
2. Here are more movies from solving the time-dependent SMOL equation for the mACHe:

<http://mccammon.ucsd.edu/smol/doc/demo/>

[mache\\_conc.mpg](#) is the ligand concentration distribution dependent on the diffusion time.

[log\\_conc.mpg](#) is the free energy flow dependent on the diffusion time.

Have fun!